

3. Werren, J.H., Windsor, D., and Guo, L. (1995). Distribution of *Wolbachia* among neotropical arthropods. *Proc. Biol. Sci.* **262**, 197–204.
4. Weinert, L.A., Araujo-Jnr, E.V., Ahmed, M.Z., and Welch, J.J. (2015). The incidence of bacterial endosymbionts in terrestrial arthropods. *Proc. Biol. Sci.* **282**, 20150249.
5. Turelli, M., Cooper, B., Richardson, K., Ginsberg, P., Peckenpaugh, B., Antelope, C., Kim, K., May, M., Abrieux, A., Wilson, D., *et al.* (2018). Rapid global spread of wRi-like *Wolbachia* across multiple *Drosophila*. *Curr. Biol.* **28**, 963–971.
6. Turelli, M., and Hoffmann, A.A. (1991). Rapid spread of an inherited incompatibility in California *Drosophila*. *Nature* **353**, 440–442.
7. Bourtzis, K., Androniki, A., Markakis, G., and Savakis, C. (1996). *Wolbachia* infection and cytoplasmic incompatibility in *Drosophila* species. *Genetics* **144**, 1063–1073.
8. Siozios, S., Cestaro, A., Kaur, R., Pertot, I., Rota-Stabelli, O., and Anfora, G. (2013). Draft genome sequence of the *Wolbachia* endosymbiont of *Drosophila suzukii*. *Genome Announc.* **1**, e00032–13.
9. Hamm, C.A., Begun, D.J., Vo, A., Smith, C.C.R., Saelao, P., Shaver, A.O., Jaenike, J., and Turelli, M. (2014). *Wolbachia* do not live by reproductive manipulation alone: infection polymorphism in *Drosophila suzukii* and *D. subpulchrella*. *Mol. Ecol.* **23**, 4871–4885.
10. Baldo, L., Hotopp, J.C.D., Jolley, K.A., Bordenstein, S.R., Biber, S.A., Choudhury, R.R., Hayashi, C., Maiden, M.C.J., Tettelin, H., and Werren, J.H. (2006). Multilocus sequence typing system for the endosymbiont *Wolbachia pipientis*. *Appl. Environ. Microbiol.* **72**, 7098–7110.
11. Richardson, M.F., Weinert, L.A., Welch, J.J., Linheiro, R.S., Magwire, M.M., Jiggins, F.M., and Bergman, C.M. (2012). Population genomics of the *Wolbachia* endosymbiont in *Drosophila melanogaster*. *PLoS Genet.* **8**, e1003129.
12. Kaur, R., Siozios, S., Miller, W.J., and Rota-Stabelli, O. (2017). Insertion sequence polymorphism and genomic rearrangements uncover hidden *Wolbachia* diversity in *Drosophila suzukii* and *D. subpulchrella*. *Sci. Rep.* **7**, 14815.
13. Daniels, S.B., Peterson, K.R., Strausburgh, L.D., Kidwell, M.G., and Chovnick, A. (1990). Evidence of horizontal transmission of the P transposable element between *Drosophila* species. *Genetics* **124**, 339–355.
14. Kofler, R., Hill, T., Nolte, V., Betancourt, A.J., and Schlotterer, C. (2015). The recent invasion of natural *Drosophila simulans* populations by the P-element. *Proc. Natl. Acad. Sci. USA* **112**, 6659–6663.
15. Houck, M.A., Clark, J.B., Peterson, K.R., and Kidwell, M.G. (1991). Possible horizontal transfer of *Drosophila* genes by the mite *Proctolaelaps regalis*. *Science* **253**, 1125–1129.
16. Ahmed, M.Z., Breinholt, J.W., and Kawahara, A.Y. (2016). Evidence for common horizontal transmission of *Wolbachia* among butterflies and moths. *BMC Evol. Biol.* **16**, 118.
17. Kriesner, P., Hoffmann, A.A., Lee, S.F., Turelli, M., and Weeks, A.R. (2013). Rapid sequential spread of two *Wolbachia* variants in *Drosophila simulans*. *PLoS Pathog.* **9**, e1003607.
18. Henry, L.M., Peccoud, J., Simon, J.C., Hadfield, J.D., Maiden, M.J., Ferrari, J., and Godfray, H.C. (2013). Horizontally transmitted symbionts and host colonization of ecological niches. *Curr. Biol.* **23**, 1713–1717.
19. Perlman, S.J., Hunter, M.S., and Zchori-Fein, E. (2006). The emerging diversity of *Rickettsia*. *Proc. Biol. Sci.* **273**, 2097–2106.
20. Hoffmann, A.A., Montgomery, B.L., Popovici, J., Iturbe-Ormaetxe, I., Johnson, P.H., Muzzi, F., Greenfield, M., Durkan, M., Leong, Y.S., Dong, Y., *et al.* (2011). Successful establishment of *Wolbachia* in *Aedes* populations to suppress dengue transmission. *Nature* **476**, 454–457.

## Social Spaces: Place Cells Represent the Locations of Others

É. Duvelle and K.J. Jeffery

Institute of Behavioural Neuroscience, Division of Psychology and Language Sciences, University College London, 26 Bedford Way, London WC1H 0AP, UK

Correspondence: [e.duvelle@ucl.ac.uk](mailto:e.duvelle@ucl.ac.uk) (É.D.), [k.jeffery@ucl.ac.uk](mailto:k.jeffery@ucl.ac.uk) (K.J.J.)

<https://doi.org/10.1016/j.cub.2018.02.017>

**How does the brain represent the location of others? Recordings in rats and bats show that, along with representing self-location in an environment, some hippocampal neurons are modulated by the position of another individual.**

As well as knowing one's own location, being able to recognize the position of others in space is a crucial skill for hunting, escaping a predator, finding a mate, or socializing. Two recent papers, by Danjo *et al.* [1] and Omer *et al.* [2], report that some cells within the hippocampus of the rat and the bat, which support self-localization, also fire in response to the location of a conspecific.

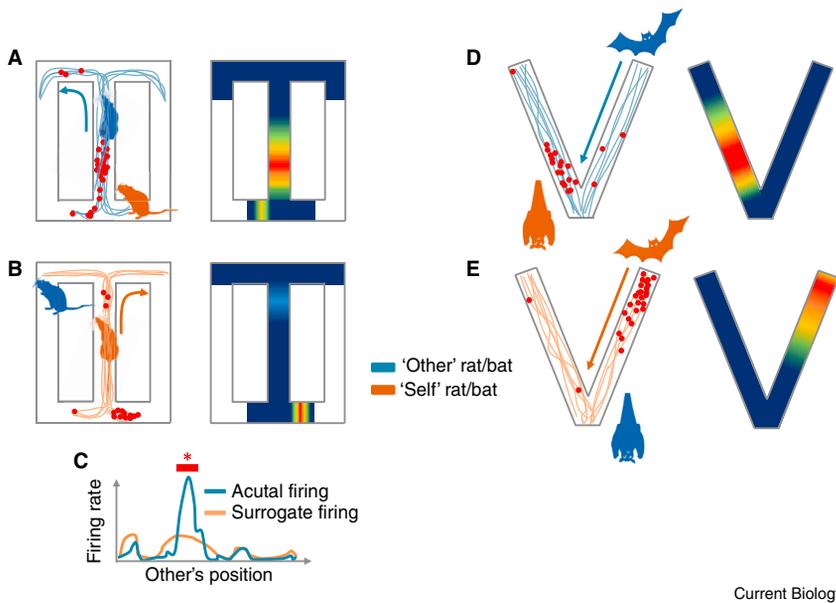
The hippocampus constructs a representation of self-location via the

activity of the spatially sensitive 'place cells' [3,4], but it has been speculated that place cells might also encode the location of other individuals. Increasing evidence suggests a role for social processing in hippocampus. For example, social stimuli induce alterations in place cell activity [5], and ventral hippocampus has been implicated in memory for individuals [6], while neuroimaging studies in humans have revealed hippocampal involvement in social rank processing [7]. Also, Mou

and Ji [8] recently found place cell reactivation in a rat observing another rat at the relevant locations. There has, however, hitherto been no clear evidence of neurons encoding the position of another individual *per se*. But now Danjo *et al.* [1] and Omer *et al.* [2] have reported such activity in the place cells of rats and bats, respectively.

Both groups [1,2] used paradigms in which an observer animal attended to the location of another 'demonstrator' animal





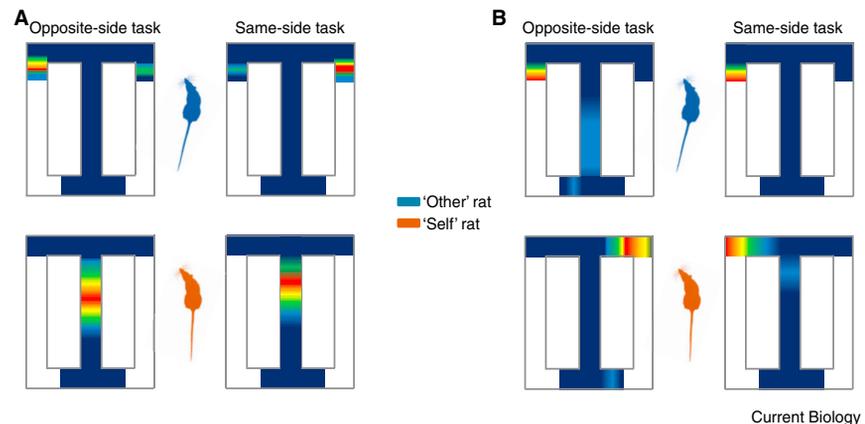
**Figure 1. Hippocampal cells encode the spatial location of other individuals.**

(A,B) The two-choice rat task [1]. The 'self' observer rat (orange) watches the 'other' demonstrator rat (blue) run up the central stem and make a choice, and then goes to the same or opposite location (depending on task rule). (A) The data plotted in the reference frame of the other. Left: spike plot of a place cell from the self rat, where spikes (red dots) are shown plotted over the other rat's trajectory (blue lines). The spike's from the self's place cells are emitted when the other rat is at particular places on its trajectory. Right: the same data plotted as a firing-rate map (red, max firing; blue, min/no firing) showing the place field in the other reference frame. (B) Left: the same data plotted in the reference frame of the self rat (trajectory, orange lines), showing activity clustered where the self rat was waiting in the right-hand start arm. Right: corresponding rate map. (C) Firing in the 'other' reference frame (blue line), compared with predicted firing constructed using the cell's firing under the hypothesis that it only depends on the self's positions (orange line). The red line and asterisk show significant 'other' firing, over and above that predicted by the self's location. (D,E) The two-choice bat task [2], illustrated as for (A,B), using in-flight data in one travel direction, showing a cell with one place field in the 'other' reference frame and a different one in the 'self' frame.

Current Biology

to solve a two-choice spatial task. In the Danjo *et al.* [1] experiment, observer rats watched a demonstrator rat take random left or right trajectories on a T-maze and then either followed the demonstrator or took the opposite route, depending on task condition. In the Omer *et al.* [2] experiment, bats watched another bat fly to food located either to the left or to the right, before flying to that same location and back. In both experiments, when cell firing was correlated with the animal's own position then most cells produced 'place fields', that is location-specific activity, in the usual way. However, a subset of the cells showed activity that was also modulated by the position in space of *another* individual (Figure 1). In the bat experiment, where trajectories went in both directions, cells were directional and usually preferred only one of the two possible trajectories, ruling out simple encoding of distance or time [9].

Were cells responding to the other animal itself or to the future trajectory of the self? Danjo *et al.* [1] decoupled self



Current Biology

**Figure 2. Two forms of beyond-self-location encoding.**

(A) A cell tuned to the self's future trajectory, independent of the other rat. Top: firing rate map in the 'other' reference frame. Bottom: same cell's rate map in the 'self' reference frame. The cell's firing in the 'other' reference frame was firing-rate-modulated (hot colors, higher rate) by the self's future goal. (B) A cell tuned to the location of the other rat, independent of the self's trajectory; reference frames as in (A). The cell always fired when the other rat was at the left goal.

stationary or following at a stereotypical distance, and so constitutes a self's place field that is modulated by the observer's position.

This type of dual encoding — modulation of a place field by factors other than the animal's own location — often occurs when the additional factor becomes a reliable and relevant descriptor of a situation. However, the finding that not just the presence, but the *location* of another animal can be one such factor is surprising and important — other animals are dynamic, and computing where they are is a complex problem. This finding then raises a number of questions. How does the observer's brain compute the other's position in the absence of the spatial stimuli, such as directional, odometric and boundary information, that contribute to self-place field formation? Does the animal care about the characteristics of the other animal (identity, social rank, etc.) or only its location? Could *two* different individuals' positions be simultaneously represented, and how would downstream

structures untangle the various signals? Is an experimenter's position represented during experiments, and how has this affected previous place cell experiments? And, most importantly, what is the function of such 'social coding' of space?

**REFERENCES**

1. Danjo, T., Toyozumi, T., and Fujisawa, S. (2018). Spatial representations of self and other in the hippocampus. *Science* 359, 213–218.
2. Omer, D.B., Maimon, S.R., Las, L., and Ulanovsky, N. (2018). Social place-cells in the bat hippocampus. *Science* 359, 218–224.
3. O'Keefe, J., and Nadel, L. (1978). *The Hippocampus as a Cognitive Map* (Oxford: Clarendon Press).
4. Marozzi, E., and Jeffery, K.J. (2012). Place, space and memory cells. *Curr. Biol.* 22, R939–R942.
5. Alexander, G.M., Farris, S., Pirone, J.R., Zheng, C., Colgin, L.L., and Dudek, S.M. (2016). Social and novel contexts modify hippocampal CA2 representations of space. *Nat. Commun.* 7, 10300.
6. Okuyama, T., Kitamura, T., Roy, D.S., Itohara, S., and Tonegawa, S. (2016). Ventral CA1

neurons store social memory. *Science* 353, 1536–1541.

7. Kumaran, D., Banino, A., Blundell, C., Hassabis, D., and Dayan, P. (2016). Computations underlying social hierarchy learning: Distinct neural mechanisms for updating and representing self-relevant information. *Neuron* 92, 1135–1147.
8. Mou, X., Ji, D., Alexander, G., Farris, S., Pirone, J., Zheng, C., Colgin, L., Dudek, S., Alme, C., Miao, C., *et al.* (2016). Social observation enhances cross-environment activation of hippocampal place cell patterns. *eLife* 5, 18428–18435.
9. MacDonald, C.J., Lepage, K.Q., Eden, U.T., and Eichenbaum, H. (2011). Hippocampal "time cells" bridge the gap in memory for discontinuous events. *Neuron* 71, 737–749.
10. Wood, E.R., Dudchenko, P.A., Robitsek, R.J., and Eichenbaum, H. (2000). Hippocampal neurons encode information about different types of memory episodes occurring in the same location. *Neuron* 27, 623–633.
11. Ferbinteanu, J., and Shapiro, M.L. (2003). Prospective and retrospective memory coding in the hippocampus. *Neuron* 40, 1227–1239.
12. Zynjuk, L., Huxter, J., Muller, R.U., and Fox, S.E. (2012). The presence of a second rat has only subtle effects on the location-specific firing of hippocampal place cells. *Hippocampus* 22, 1405–1416.

# Infection Models: Novel Potato Blight-like Pathogens in Worms

Hinrich Schulenburg

Zoological Institute, Christian-Albrechts-University Kiel, 24098 Kiel, Germany; and Max-Planck Institute for Evolutionary Biology, 24306 Plön, Germany

Correspondence: [hschulenburg@zoologie.uni-kiel.de](mailto:hschulenburg@zoologie.uni-kiel.de)

<https://doi.org/10.1016/j.cub.2018.02.018>

Oomycetes are best known as plant pathogens, causing for example potato blight. Other oomycetes are deadly yet less well studied pathogens of animals including humans. Osman and colleagues now present the nematode *C. elegans* as a new, genetically tractable host model that should enhance our general understanding of oomycete infections.

Potato blight has had devastating effects on potato harvest, repeatedly causing serious famines in human populations. The most infamous example is the Great Irish Famine in the middle of the 19th century, in which potato blight accounted for a death toll of more than one million, leading another two million to emigrate to the United States. Potato blight is caused

by a pathogen with a fateful name, *Phytophthora infestans*, meaning the plant-ruining infestation. This pathogen belongs to an unusual group of organisms, the oomycetes, that were long thought to be fungi, but which are in fact more closely related to brown algae and diatoms [1]. *Phytophthora* and other oomycetes are still considered to pose a

serious threat to economically relevant plants and are thus of particular interest to agricultural research [2]. It is less well known that oomycetes can also infect animals and cause disease in humans. One example includes pythiosis caused by *Pythium insidiosum*, which is now considered an emerging human pathogen with high morbidity and significant

